### *JP XVII* (2016)

## JP XVII (2016)

### Shakuyakukanzoto Extract

芍薬甘草湯エキス

Shakuyakukanzoto Extract contains not less than 50 mg and not more than 150 mg of paeoniflorin  $(C_{23}H_{28}O_{11}: 480.46)$ , and not less than 50 mg and not more than 150 mg of glycyrrhizic acid  $(C_{42}H_{62}O_{16}: 822.93)$ , per extract prepared with the amount specified in the Method of preparation.

Method of preparation		
	1)	2)
Peony Root	6 g	5 g
Glycyrrhiza	6 g	5 g

Prepare a dry extract or viscous extract as directed under Extracts, according to the prescription 1) or 2), using the crude drugs shown above.

**Description** Shakuyakukanzoto Extract occurs as a light brown powder or brown viscous extract. It has slightly an odor, and a sweet taste.

**Identification** (1) Shake 0.5 g of dry extract (or 1.5 g of the viscous extract) with 10 mL of water, then add 10 mL of 1-butanol, shake, centrifuge, and use the supernatant liquid as the sample solution. Separately, dissolve 1 mg of Paeoniflorin RS or paeoniflorin for thin-layer chromatography in 1 mL of methanol, and use this solution as the standard solution. Perform the test with these solutions as directed under Thin-layer Chromatography  $\langle 2.03 \rangle$ . Spot 5  $\mu$ L each of the sample solution and the standard solution on a plate of silica gel for thin-layer chromatography. Develop the plate with a mixture of ethyl acetate, methanol and water (20:3:2) to a distance of about 10 cm, and air-dry the plate. Spray evenly 4-methoxybenzaldehyde-sulfuric acid TS on the plate, and heat at 105°C for 5 minutes: one of the spot among the several spots obtained from the sample solution has the same color tone and Rf value with the purple spot obtained from the standard solution (Peony Root).

(2) Shake 0.5 g of dry extract (or 1.5 g of the viscous extract) with 10 mL of water, then add 10 mL of 1-butanol, shake, centrifuge, and use the supernatant liquid as the sample solution. Separately, dissolve 1 mg of liquiritin for thinlayer chromatography in 1 mL of methanol, and use this solution as the standard solution. Perform the test with these solutions as directed under Thin-layer Chromatography <2.03>. Spot 5  $\mu$ L each of the sample solution and the standard solution on a plate of silica gel for thin-layer chromatography. Develop the plate with a mixture of ethyl acetate, methanol and water (20:3:2) to a distance of about 10 cm, and air-dry the plate. Spray evenly dilute sulfuric acid on the plate, and heat at 105°C for 5 minutes: one of the spot among the several spots obtained from the sample solution has the same color tone and Rf value with the yellow-brown spot obtained from the standard solution (Glycyrrhiza).

**Purity (1)** Heavy metals  $\langle 1.07 \rangle$ —Prepare the test solution with 1.0 g of the dry extract (or an amount of the viscous extract, equivalent to 1.0 g of dried substance) as directed under Extracts (4), and perform the test (not more than 30 ppm).

(2) Arsenic  $\langle 1.11 \rangle$ —Prepare the test solution with 1.0 g of the dry extract (or an amount of the viscous extract, equivalent to 1.0 g of dried substance) according to Method 3, and perform the test (not more than 2 ppm).

Loss on drying  $\langle 2.41 \rangle$  The dry extract: Not more than 8.0% (1 g, 105°C, 5 hours).

The viscous extract: Not more than 66.7% (1 g,  $105^{\circ}$ C, 5 hours).

**Total ash** <5.01> Not more than 9.0%, calculated on the dried basis.

Assay (1) Paeoniflorin—Weigh accurately about 0.2 g of the dry extract (or an amount of the viscous extract, equivalent to 0.2 g of dried substance), add exactly 50 mL of diluted methanol (1 in 2), shake for 15 minutes, filter, and use the filtrate as the sample solution. Separately, weigh accurately about 10 mg of Paeoniflorin RS (separately determine the water  $\langle 2.48 \rangle$  by coulometric titration, using 10 mg), dissolve in diluted methanol (1 in 2) to make exactly 100 mL, and use this solution as the standard solution. Perform the test with exactly 10  $\mu$ L each of the sample solution and standard solution as directed under Liquid Chromatography  $\langle 2.01 \rangle$  according to the following conditions, and determine the peak areas,  $A_{\rm T}$  and  $A_{\rm S}$ , of paeoniflorin in each solution.

Amount (mg) of paeoniflorin (C<sub>23</sub>H<sub>28</sub>O<sub>11</sub>)  
= 
$$M_{\rm S} \times A_{\rm T}/A_{\rm S} \times 1/2$$

 $M_{\rm S}$ : Amount (mg) of Paeoniflorin RS taken, calculated on the anhydrous basis

### Operating conditions—

Detector: An ultraviolet absorption photometer (wave-length: 232 nm).

Column: A stainless steel column 4.6 mm in inside diameter and 15 cm in length, packed with octadecylsilanized silica gel for liquid chromatography (5  $\mu$ m in particle diameter).

Column temperature: A constant temperature of about  $20^{\circ}$ C.

Mobile phase: A mixture of water, acetonitrile and phosphoric acid (850:150:1).

Flow rate: 1.0 mL per minute (the retention time of paeoniflorin is about 9 minutes).

System suitability—

System performance: Dissolve 1 mg each of Paeoniflorin RS and albiflorin in diluted methanol (1 in 2) to make 10 mL. When the procedure is run with  $10 \,\mu$ L of this solution under the above operating conditions, albiflorin and paeoniflorin are eluted in this order with the resolution between these peaks being not less than 2.5.

System repeatability: When the test is repeated 6 times with  $10 \,\mu\text{L}$  of the standard solution under the above operating conditions, the relative standard deviation of the peak area of paeoniflorin is not more than 1.5%.

(2) Glycyrrhizic acid—Weigh accurately about 0.2 g of the dry extract (or an amount of the viscous extract, equivalent to 0.2 g of dried substance), add exactly 50 mL of diluted methanol (1 in 2), shake for 15 minutes, filter, and use the filtrate as the sample solution. Separately, weigh accurately about 10 mg of Glycyrrhizic Acid RS (separately determine the water  $\langle 2.48 \rangle$  by coulometric titration, using 10 mg), dissolve in diluted methanol (1 in 2) to make exactly 100 mL, and use this solution as the standard solution. Perform the test with exactly 10  $\mu$ L each of the sample solution and standard solution as directed under Liquid Chromatography  $\langle 2.01 \rangle$  according to the following conditions, and determine the peak areas,  $A_{\rm T}$  and  $A_{\rm S}$ , of glycyrrhizic acid in each solution.

Amount (mg) of glycyrrhizic acid ( $C_{42}H_{62}O_{16}$ ) =  $M_S \times A_T/A_S \times 1/2$ 

 $M_{\rm S}$ : Amount (mg) of Glycyrrhizic Acid RS taken, calculated on the anhydrous basis

### Operating conditions—

Detector: An ultraviolet absorption photometer (wavelength: 254 nm).

Column: A stainless steel column 4.6 mm in inside diameter and 15 cm in length, packed with octadecylsilanized silica gel for liquid chromatography (5  $\mu$ m in particle diameter).

Column temperature: A constant temperature of about 40°C.

Mobile phase: A mixture of diluted acetic acid (31) (1 in 15) and acetonitrile (13:7).

Flow rate: 1.0 mL per minute (the retention time of glycyr-rhizic acid is about 12 minutes).

### System suitability-

System performance: When the procedure is run with 10  $\mu$ L of the standard solution under the above operating conditions, the number of theoretical plates and the symmetry

factor of the peak of glycyrrhizic acid are not less than 5000 and not more than 1.5, respectively.

System repeatability: When the test is repeated 6 times with  $10 \,\mu\text{L}$  of the standard solution under the above operating conditions, the relative standard deviation of the peak area of glycyrrhizic acid is not more than 1.5%.

Containers and storage Containers—Tight containers.

## *JP XVII* (2016)

# *JP XVII* (2016)

